

**INTENDED USE**

The Thyroglobulin Auto-Antibody Test Kit is used for detection of Thyroglobulin Auto-Antibody (TgAA) in canine plasma or serum.

ANALYTICAL PRINCIPLE

Canine Thyroglobulin IgG-specific antibody in diluted samples is allowed to bind to microwell-bound canine thyroglobulin antigen. After washing unbound materials, HRP-conjugate is allowed to bind to the Thyroglobulin IgG antibody-antigen complex. Unbound HRP-conjugate is washed away and TMB is allowed to react with bound HRP-conjugate. The reaction is stopped and the microwell is read. The intensity of the color produced in the HRP-TMB reaction is proportional to the amount of IgG-specific antibody in the sample.

SPECIMEN REQUIREMENTS

Serum and EDTA plasma samples are acceptable. Avoid repetitive freezing and thawing of samples.

REAGENTS**Precautions & Safety Notes**

- WEAR LATEX GLOVES, FACE SHIELDS AND A LAB COAT WHEN HANDLING SPECIMENS AND OTHER HAZARDOUS REAGENTS
- FOR *IN VITRO* USE, POTENTIAL BIOHAZARDOUS MATERIAL. HANDLE ASSAY REAGENTS AS IF CAPABLE OF TRANSMITTING AN INFECTIOUS AGENT.
- The Sample Diluent, Positive Control, Negative Control, Cutoff Calibrator, Wash Buffer, and HRP-Conjugate contain 0.1% ProClin 150. Avoid contact of these reagents with skin or eyes.

Kit Components

ID	Reagent	Part Number	Quantity
	Canine Thyroglobulin Auto-Antibody Test Kit	CTGA-1000	96 Test
SD	Sample Diluent	SDFEA-1003	2 x 50 ml
A	Cutoff Calibrator See QC Certificate for value	CTGA-1001	1.5 ml
B	Positive Control See QC Certificate for value	CTGA-1002	1.5 ml
C	Microwell Plate	CTGA-1003	96 Well Plate
D	HRP Conjugate	CTGA-1004	10 ml
WB	20X Wash Buffer Concentrate	WBFEA-1001	50 ml
TS	TMB Solution Keep away from light	TMBS-1001	10 ml
SS	Stop Solution	SSFEA-1001	10 ml

MATERIALS

Materials needed but not supplied:

- Variable Pipettors and Tips
- Stir Bar & Stirrer
- 12x75 mm disposable borosilicate glass culture tubes
- Test Tube Rack, polypropylene
- Vortexer
- Microwell Plate Film Sealer
- ELISA Plate Washer

- ELISA Plate Reader
- Refrigerator (for kit storage)
- Laboratory Tape

PROCEDURE PRECAUTIONS

- Use clean instruments & equipments.
- Unused microwells should be sealed tightly using laboratory tape in the foil pouch until further use.
- Handle microwells with care.
- Minimize air bubbles in microwells.
- Microwells should not contain liquid after the washing steps. To ensure that liquids are removed from microwells after washing steps, microwells can be tapped against clean paper towels.

REAGENT PREPARATION**1X Wash Buffer Preparation**

1. Add 1 unit of volume of 20X Wash Buffer Concentrate (**WB**) to 19 units of volume of DI water. For example, add **50 ml** 20X Wash Buffer Concentrate (**WB**) to **950 ml** of DI water.
2. Mix the Wash Buffer Preparation well.

PROCEDURE

1. Dilute samples by **1:100** by adding **10 µl** of samples to **990 µl** of Sample Diluent (**SD**) in a glass tube.
2. Vortex the tubes.
3. Dispense **100 µl** of the diluted samples, ready to use Cutoff Calibrator (**A**), and ready to use Positive Control (**B**) from step 1 into designated wells of the Microwell Plate (**C**)
4. Cover the wells.
5. Incubate the wells for **30 minutes** at **room temperature**.
6. Wash each well **3 times** with **300 µl** of **1X Wash Buffer Preparation**. Tap the microwells against clean paper towels to ensure that no liquid remains.
7. Dispense **100 µl** of HRP Conjugate (**D**) to each well.
8. Cover the wells.
9. Incubate the wells for **15 minutes** at **room temperature**.
10. Wash each well **3 times** with **300 µl** of **1X Wash Buffer Preparation**. Tap the microwells against clean paper towels to ensure no liquid remains.
11. Dispense **100 µl** of TMB Solution (**TS**) to each well.
12. Cover the wells.
13. Incubate the wells for **15 minutes** at **room temperature** and **protect from light**.
14. Dispense **100 µl** of Stop Solution (**SS**) to each well.
15. Read the wells in a microwell reader at **450 nm**.

PROCEDURAL NOTES

- Evaluation of samples should be based on the Sample to Negative Ratio.

EXPECTED VALUES**Normal Sample Study**

One-hundred and sixty (160) normal dog samples were assayed with IVD Technologies' Canine Thyroglobulin Auto-Antibody Test Kit. The O.D. ranged from 0.07 to 0.500.



Sample to Negative Ratio Calculation

Sample to Negative Ratio

= O.D. Sample / O.D. Cutoff Calibrator

- The Sample to Negative Ratio is equal to the O.D. of the Sample over the O.D. of the Cutoff Calibrator.

Sample to Negative Ratio Table

< 0.900	Negative
0.900 – 1.20	Borderline
> 1.20	Positive

CORRELATION ANALYSIS

A total of 180 canine samples were assayed for thyroglobulin antibody using IVD Technologies' Canine Thyroglobulin Auto-Antibody ELISA Test Kit and a Reference Laboratory's Canine Thyroglobulin Auto-Antibody ELISA Test Kit. Fifteen (15) samples were found to be positive by IVD Technologies, while 14 samples were found to be positive by the Reference Laboratory. One-hundred and sixty-five (165) samples were found to be negative by IVD Technologies, while 166 samples were found to be negative by the Reference Laboratory.

**Correlation Analysis Table
Reference Laboratory
CTgAA ELISA**

IVD Technologies CTgAA ELISA	POSITIVE	NEGATIVE	Total
POSITIVE	13	2	15
NEGATIVE	1	164	165
Total	14	166	180

Assay Sensitivity and Specificity

Based on the correlation table, IVD Technologies' Canine Thyroglobulin Auto-Antibody ELISA Kit has a sensitivity of 92.9% and a specificity of 98.8%.

Sensitivity	92.9%
Specificity	98.8%

REPRODUCIBILITY

Intra-Assay

Three (3) samples were assayed in eight (8) replicates in a single assay. The mean, standard deviation, and CVs are listed in the table below.

Sample ID	1	2	3
Replicates	8	8	8
Mean	0.25	0.12	1.87
SD	0.0250	0.0030	0.107
CV	10	3.1	5.7

Inter-Assay

Three (3) samples were tested in six (6) assays. The mean, standard deviation, and CVs are listed in the table below

Sample ID	A	B	C
Assays	6	6	6
Mean	0.1	0.34	1.09
SD	0.001	0	0.046
CV	1.03	1.32	4.27

REFERENCES

Beale, K. M., R. E. Halliwell, and C. L. Chen, 1990: Prevalence of antithyroglobulin antibodies detected by enzyme-linked immunosorbent assay of canine serum. *J. Am. Vet. Med. Assoc.* 196, 745–748.

Beale, K. M., 1991: Canine immune mediated thyroiditis: the role of autoantibodies in diagnosis. *Vet. Med. Rep.* 3, 123–130.

Dent, A. H., 2001: Conjugation methods. In: Wild, D. (ed.) *The Immunoassay Handbook*, 2nd edn, pp. 211–228. Nature Publishing Group, London and Basingstoke.

Dixon, R. M., and C. T. Mooney, 1999: Canine serum thyroglobulin autoantibodies in health, hypothyroidism and non-thyroidal illness. *Res. Vet. Sci.* 66, 243–246.

Ericsson, U. B., I. Larsson, and J. I. Thorell, 1984: Purification and storage of thyroglobulin. Two important factors influencing the radioimmunoassay for thyroglobulin. *Scand. J. Clin. Lab. Invest.* 44, 477–485.

Rajatanavin, R., S. L. Fang, S. Pino, P. Laurberg, L. E. Braverman, M. Smith, and L. P. Bullock, 1989: Thyroid hormone antibodies and Hashimoto's thyroiditis in mongrel dogs. *Endocrinology* 124, 2535–2540.

Manufacturer Made in the USA
IVD Technologies 2002 S. Grand Avenue, Suite A Santa Ana, CA 92705 USA Tel: 1(714)549-5050 Fax: 1(714)549-5055 info@ivdtechnologies.com www.ivdtechnologies.com